

# NREL-Amoco CRADA Phase 3

## Bench Scale Report 1.7

### Batch SSCF of Pretreated Corn Fiber with LNHST2

**Project Title:** Amoco-NREL CRADA with corn fiber

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#### Objectives

Several objectives were pursued in this study: (1) **Determine** if a solids loading of 20% corn fiber **has an** effect on the fermentation performance of the yeast strain LNHST2 compared to a 10% **solids** loading; (2) Ascertain the time it takes in batch mode to **consume** 90% of the soluble xylose; (3) Monitor the course of the soluble sugars, xylans, **and** glucans in the reactor **and** determine the ethanol **and** by-product **yields**; and (5) With these data, close the carbon balance around the **SSCF** for each solids loading.

#### Background

This experiment **is** designed to support the **Task 3 PDU** activities, which will examine the operability of the pilot plant with APR-pretreated corn fiber solids, **using** the organism LNHST2. Results **obtained** at the bench scale can then be compared to results obtained at the **PDU** to examine the **ability** to scale up lab data.

#### Materials and Methods

##### *Inoculum*

Inoculum **was** generated in YEPD (1% w/v yeast extract, 2% w/v peptone **and** 2% w/v glucose, pH 5.0) in two stages at 30°C **and** 150 rpm. The first stage, consisting of 50 mL of YEPD in a 250 mL baffled Erlenmeyer **flask**, was inoculated **with** a colony from **a** YEPX plate (1% w/v yeast extract, 2% w/v peptone **and** 2% w/v glucose, pH 5) **and** incubated for 14.5 hours. A 10% w/v inoculum was added to two **500** mL baffled Erlenmeyer **flasks** containing 130 mL of YEPD each. The **flasks** were incubated for only 6 hours; by that time, a majority of the glucose was consumed, but not **all** (exponential growth stage). The contents of **both flasks were** combined, mixed **and** split to ensure the same **inoculum** for each fermentor.

##### *Enzyme and nutrients*

Cellulase enzyme was used with an activity determined at NREL to be 80 IFPU/mL. Based on this activity and **an** assumed 25% cellulose content in **raw** corn fiber, **an** amount of cellulase **was** added to each fermentor corresponding to 10 IFPU/g cellulose. In

addition to cellulase, glucoamylase **was** acquired from **Enzyme** Development Corporation with a reported activity of 285 IU/mL. The enzyme was added at a level of 2 IU/g of starch based on a **20%** starch content assumption in raw corn fiber. Each enzyme preparation **was** filter sterilized through 0.2  $\mu$ m filter before **being** added to the fermentors.

Corn Steep Liquor (CSL) (1% w/v) **was** added **as** a nutrient source to each fermentor. To prepare the solution, the pH of a 50% w/w dilution of the purchased CSL **was** first adjusted to 5.0 with sodium hydroxide **pellets** and autoclaved

### *Corn fiber*

Corn fiber (**CF**) **was** obtained from the APR.

As the pH of the **CF** is very low, the pH needs to **be** adjusted to 5.0 for the fermentation. In order to do this, 1500 grams of material was weighed out, 210 mL of water was added to facilitate **mixing**, and 49 mL of a 10M NaOH **solution** was **added** to bring the pH to 5.0. Based on the added liquids, the percent **solids** was recalculated (25.59%) and verified by measuring the solids level with **an** IR balance (**25.50%**).

### *Fermentations*

The amount **of** CF necessary to give 10% and **20% solids** in a 1.2-L final volume **was** weighed **and** placed into the two BioFlo III fermentors with a known amount of **additional** water **and** autoclaved **for** one hour. **Each** fermentor **was** inoculated with the appropriate amount of inoculum, CSL, enzymes, and enough water to **bring** the solids level to the appropriate level. **Each** fermentor **was** operated at 30°C, 150 **rpm**, and pH 5.0, which was controlled **with** the addition of 3 M sodium hydroxide.

### *Sampling and Analysis*

**Initial** and final samples were obtained and a total compositional analysis **was** performed on the **solid and** liquid fractions. Samples were taken every **24** hours and the liquid fraction was analyzed for cellobiose, apparent xylitol, lactic acid, glycerol, acetic acid, HMF, **and** furfural, as well **as** monomeric **and** oligomeric sugars (glucose, xylose, galactose, arabinose, **and** mannose). Colony forming **units** (CFUs) were also performed on each sample to monitor the cell population.

## **Results and Discussions**

### *Effect of Solids Loading on Fermentation Performance*

**Based** on glucose and xylose **utilization**, ethanol **production**, and cell counts, **there is** no discernible effect **from** the **high** solids loading (20% extruded corn fiber) on the fermentation performance of LNHST2. The initial monomeric **and** soluble glucose concentrations in the 20% **solids SSCF** **are** twice those of the 10% solids **SSCF**, as seen in Table 1. Within **22** hours **of** fermentation, 86-90% of the glucose **was** consumed in both the 10% and **20% SSCF**. **By normalizing** the glucose results, it is apparent that a similar percentage of glucose is consumed **regardless** of the **solids** loading (Table 1).

**Table 1:** Soluble glucose utilization in 10% and 20% corn fiber solids SSCF

Elapsed Time (h)	20% solids	10% Solids	20% solids	10% Solids
	Soluble Glucose	Soluble Glucose (g/L)	Normalized Glucose (%)	Normalized Glucose (%)
0	57.84	30.13	100.0	100.0
22	8.10	3.02	14.0	10.0
46	<b>5.69</b>	<b>2.55</b>	9.84	8.46
<b>68.5</b>	5.45	2.38	9.42	7.90
91.5	5.36	2.52	9.27	8.37
120	5.37	23 1	9.28	7.67

Table 2 and Figures 1 and 2 show the soluble xylose (monomeric and oligomeric xylose) concentrations. In contrast to glucose, xylose consumption in the two reactors is not exactly the same. It is not known to what extent analytical errors are responsible for the inconsistency, but it should be noted that the initial xylose concentration levels are quite low, making the analysis less accurate. The normalized soluble xylose values show a slight increase in the xylose consumption at the 10% solids level over the 20% solids level (Table 2). Interestingly, an increase in oligomeric xylose is observed between zero and 22 hours at both solids levels (Figures 1 and 2). This may be due to an analytical error on the time zero sample, since there is no evidence of xylanase activity within the cellulase enzyme complex. Beyond the first 22 hours of the SSCF, the oligomeric xylose concentration remained unchanged

Unfortunately, 90% xylose consumption (a PDU objective) was not achieved within the duration of the experiment (120 h). Only 64.6% of the monomeric xylose was consumed in the 20% SSCF and 85.1% was consumed in the 10% SSCF in 120 h. Based on the utilization of monomeric xylose, the consumption rate at the 20% level was 0.041 g/L-h, only slightly lower than the 0.053 g/L-h at 10% solids.

**Table 2:** Soluble xylose utilization in 10% and 20% corn fiber solids SSCF

Elapsed Time (h)	20% solids		10% Solids		20% solids	10% Solids
	Total Soluble Xylose (g/L)	Monomeric Xylose (g/L)	Total Soluble Xylose (g/L)	Monomeric Xylose (g/L)	Normalized Monomeric Xylose	Normalized Monomeric Xylose (%)
0	18.00	3.93	10.39	2.08	100	100
22	25.83	3.24	11.73	.9 1	82.4	43.8
46	24.80	2.05	11.34	.5	52.2	24.0
68.5	23.89	1.56	11.23	.37	39.7	17.8
91.5	24.27	1.39	11.31	.3 1	<b>35.4</b>	14.9
120	24.81	1.39	10.96	.3 1	35.4	14.9

Another parameter that can be examined to determine if the solids loading has an effect on the fermentation performance is ethanol production. Unfortunately, problems were encountered with the analysis of the initial solids samples. Without this piece of data, theoretical ethanol yields cannot be calculated. Nevertheless, ethanol yields were calculated based on the initial solids loadings and expressed as grams of ethanol produced per gram of initial solids. The data show that there was no substantial difference in the ethanol production at 20% solids compared to 10% solids (Table 3). The yield profile was similar in the two reactors. In both vessels, the ethanol concentration reached its peak value within approximately 70 hours of SSCF.

**Table 3:** Ethanol Production in 10% and 20% extruded corn fiber solids SSCF

Elapsed Time (h)	20%solids Residual Ethanol Concentration (g/L)	10%Solids Residual Ethanol Concentration (g/L)	20%solids Ethanol Yield (g ethanol/ g solids)	10%Solids Ethanol Yield (g ethanol/ g solids)
0	1.08	1.01		
22	32.07	16.69	0.148	0.157
46	34.57	17.61	0.167	0.166
68.5	38.46	18.50	0.187	0.175
91.5	37.85	18.70	0.184	0.177
120	37.05	18.54	0.180	0.175

In addition to sugar utilization and ethanol production, cell concentration was also monitored in both fermentations. The cell concentrations were similar in the two reactors:  $9.5 \times 10^7$  cells/mL in the 20% solids and slightly greater at  $1.4 \times 10^8$  cells/mL in the 10% solids after 22 hours of growth. A decline in cell growth was observed at both solids levels beyond 70 hours of SSCF, perhaps as a result of starvation from monomeric sugars.

#### *Fermentation Duration*

Within 22 hours, a majority of the soluble glucose is consumed and a majority of the ethanol is produced at both solids levels (Figure 3). Only a small amount of the monomeric xylose and none of the oligomeric xylose is utilized within that period. Over the rest of the fermentation, more monomeric xylose is converted to ethanol (Figure 3).

#### *Yields, By-products, and Sugar Conversions*

Yields and by-products were calculated based on the total glucose and xylose present at each solids loading. Glucan (in equivalent glucose) conversion in the 10% solids SSCF was 87.64% compared to 84.37% at 20% solids (see Table 4). This low conversion is due to the inefficient conversion of cellulose and starch to glucose during pretreatment. This is also evidenced in the xylose conversions where only 15.86% and 14.23% of the total available xylan (in equivalent xylose) is converted in the 10% and 20% solids SSCFs respectively. The lack of monomeric xylose production during pretreatment is even more severe than glucose as the enzyme preparation does not work effectively against oligomeric xylose and xylose in the solids.

**Table 4:** Sugar Conversions and Ethanol Yields based on Total Glucose and Xylose

<b>Initial Solids Loading</b>	10%	20%
Glucose Conversion(%)	<b>87.64</b>	<b>84.37</b>
Xylose Conversion (%)	15.86	14.23
Ethanol Process Yield (% theoretical)	60.13	50.61
Ethanol Metabolic Yield (% theoretical)	91.52	82.63

The ethanol process yield (based on total glucose and xylose) at 10% solids was 60.13% of theoretical **and** lower at 50.61% of theoretical at 20% solids (see Table 4). The ethanol metabolic yield at 10% solids was also higher (91.52%) than at **20%** solids (82.63%). This result could indicate ~~that~~ there is an effect of the solids loading on the fermentation performance. However, when the carbon balance is examined (see Table 5), the closure at the 10% solids level is well over 100% indicating that the obtained **yield** values could be inflated (analytical error). In addition, the 20% solids SSCF closure is under 100% indicating that the yields **at** the 20% solids loading could **be** deflated. One reason for the **disparity** in the carbon balance closures is ~~that~~ the value **used** to calculate the glucose concentration in the solids **is** very sensitive to the percent insoluble solids, which **is** calculated and not determined.

The major by-products observed were glycerol at 0.025 g/g consumed sugar and 0.029 g/g consumed sugar and xylitol at 0.052 g/g consumed sugar **and** 0.025 g/g consumed sugar in the 10% and **20%** solids SSCF respectively (see Table 5).

**Table 5:** Product Distribution for the 10% and 20% Solids SSCFs (expressed **as grams** of product per 100grams of consumed glucose and xylose)

Product	10%solids Loading	20% solids Loading
Ethanol	46.77	42.22
<del>Carbon</del> Dioxide	47.29	41.55
<del>Cell</del> Mass	5.02	5.08
Glycerol	<b>2.54</b>	2.91
Acetic Acid	0.26	0.93
Succinic Acid	<b>1.64</b>	0.38
Xylitol	5.02	2.50
Total	<b>108.54</b>	95.58

## Conclusions

The time it took for the majority of the monomeric glucose to be consumed **was** just 22 hours. Xylose **was** taken up at a slower rate. Based on the available data, there **is** no discernible effect of the solids loading on the fermentation performance of LNHST2. The ~~cell~~ concentration in the 20% solids fermentation was similar to ~~that~~ in the 10% solids fermentation, **and** 3.8% w/v ethanol was produced **from** the 20% solids, double the 1.9% produced from the 10% solids. The 3.8% w/v ethanol and 1.9% w/v ethanol produced in the 20% and 10% solids fermentations, respectively,

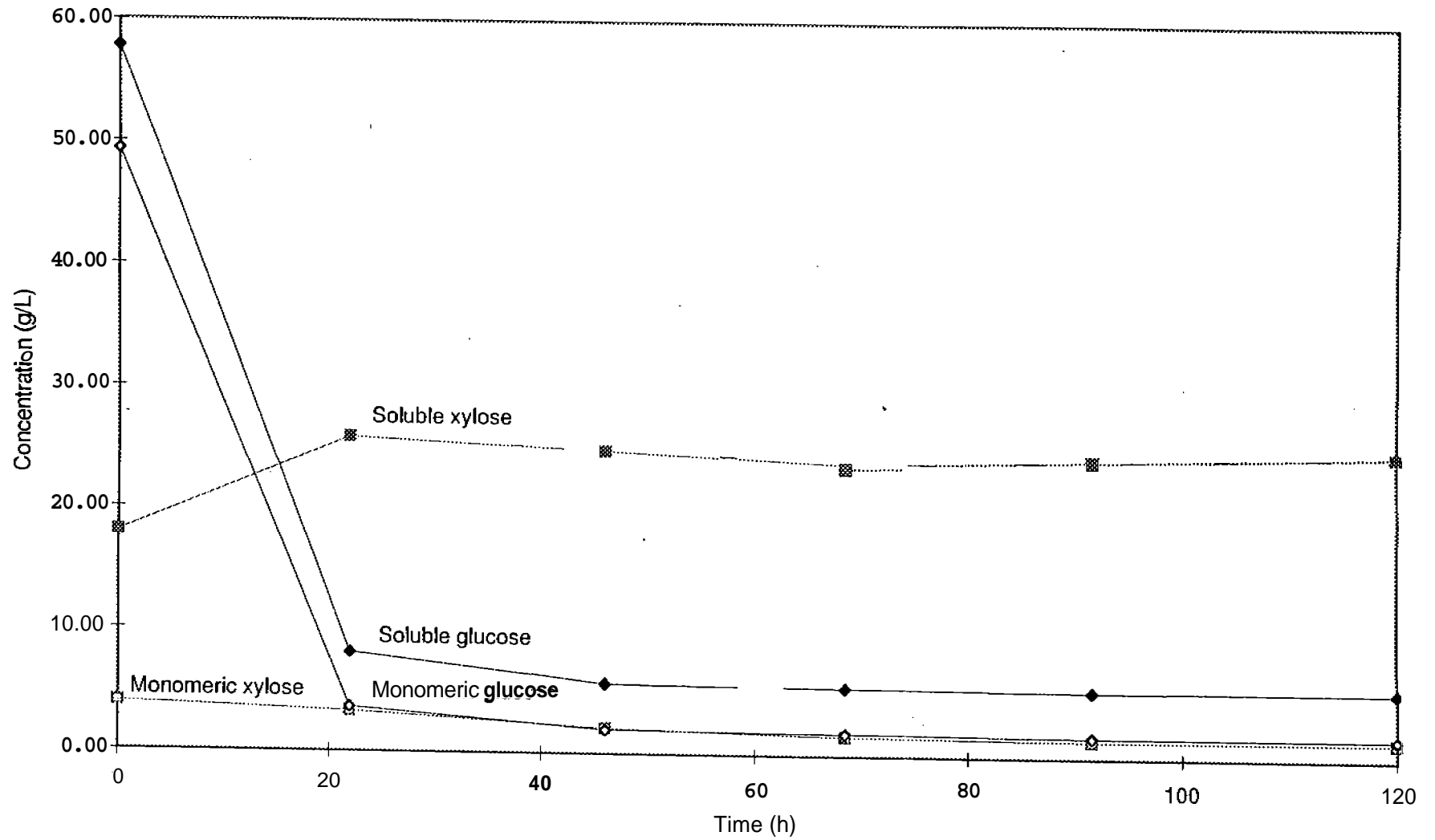
exceed the theoretical yields based on the consumed soluble sugars. The **solids data** will **allow** us to calculate the true yield coefficients.

Another point of interest in **this** experiment is the **lack** of conversion of oligomeric xylose to monomeric xylose. In the absence of xylanase activity in the cellulase enzyme preparation, it seems clear **that** the pretreatment procedure needs to convert more **xylans** to monomeric **xylose** in order to capitalize on the xylose-fermenting capability of the **yeast strain** LNHST2.

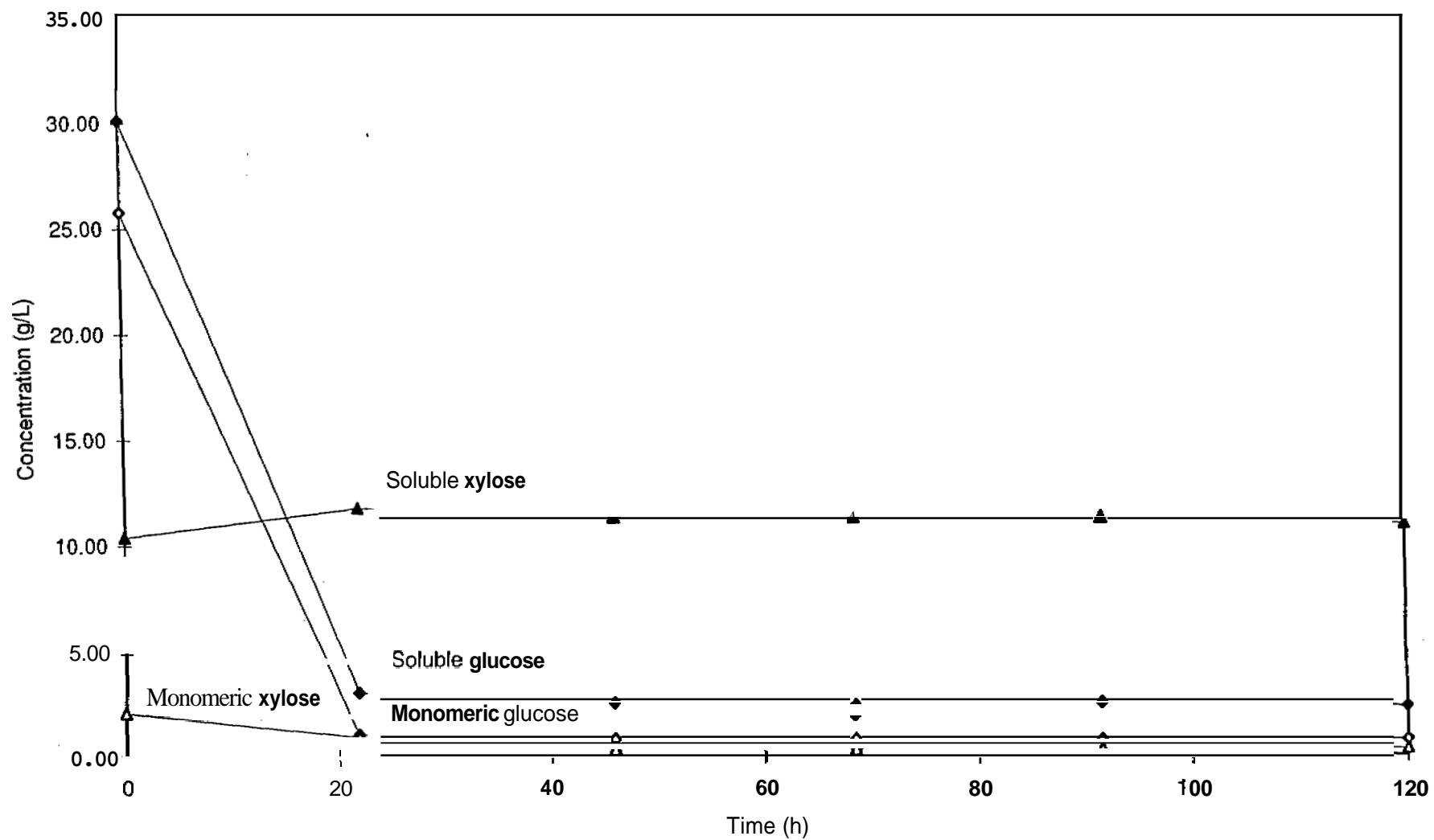
The insoluble **solids** content **of** pretreated biomass **is** critical in calculating the glucose **and** xylose present in biomass. Without a reliable value **of** insoluble **solids**, the value of the yield information **is diminished**. We **should** strive in the **future** to **come** up with a better method for determining **this** value.

Figure 1: Soluble Sugar Profile in 20% Corn Fiber Solids SSCF with

LNHST2

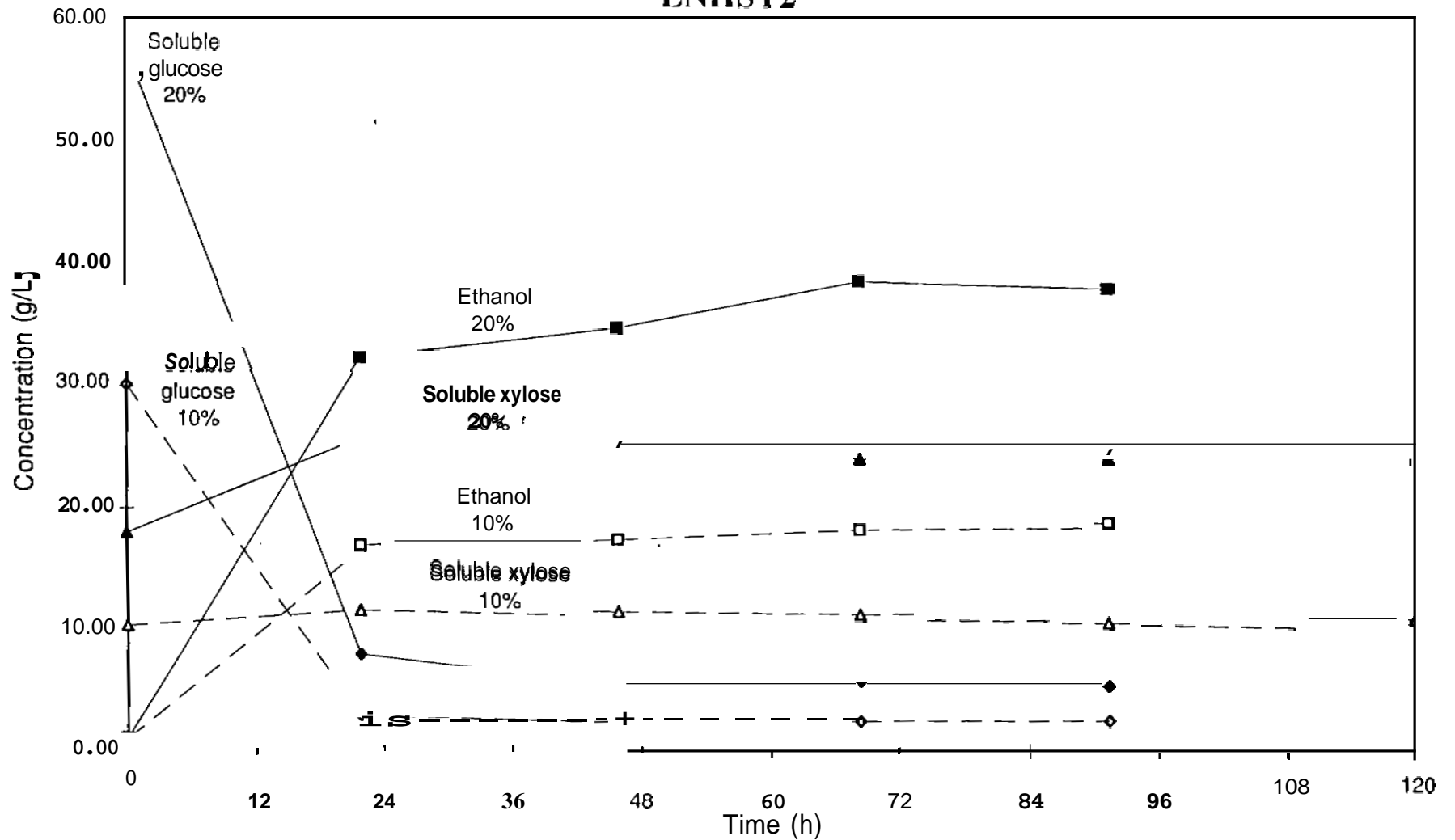


**Figure 2: Soluble Sugar Profile in 10% Corn Fiber Solids SSCF with LNHST2**





**Figure 3: Comparison of 10% and 20% Corn Fiber Solids SSCF with LNHST2**



# Raw Data

## 20% Initial Solids Level - original CAT task data

		YSI		YSI				Hauser				Hauser				Hauser		
Elapsed time (h)	Solids (%)	Glucose (g/L)	Glucose (g/L)	Ethanol (g/L)	Ethanol (g/L)	Xylose (g/L)	Cellobiose (g/L)	Monomeric Glucose (g/L)	Soluble Glucose <sup>1</sup> (g/L)	Oligomeric Glucose <sup>1,2</sup> (g/L)	Monomeric Xylose (g/L)	Soluble Xylose (g/L)	Oligomeric Xylose <sup>2</sup> (g/L)	Galactose (g/L)	Total Galactose (g/L)	Arabinose (g/L)	Total Arabinose (g/L)	
0	19.40		49.65	0.97	1.08	7.44	5.25	49.32	57.84	8.52	3.93	18.00	14.07	1.05	4.20	12.18	20.22	
22	15.78	0.36	0.90	34.10	32.07	5.92	0.00	3.60	8.10	4.50	3.24	25.83	22.59	0.84	5.01	12.00	21.42	
46		0.19	0.00	37.20	34.57	3.33	0.00	1.94	5.69	3.75	2.05	24.80	22.75	0.87	5.17	11.90	20.52	
68.5	15.20	0.19	0.00	37.10	38.46	2.93	0.00	1.80	5.45	3.65	1.56	23.89	22.33	0.85	4.93	11.61	19.66	
91.5	15.29	0.16	0.00	38.29	37.85	2.83	0.00	1.69	5.36	3.67	1.39	24.27	22.88	0.89	5.05	11.36	19.95	
120	14.91	0.16		37.05			0.00	1.69	5.37	3.68	1.39	24.81	23.42	0.96	5.21	11.36	20.00	
Updated CAT task values																		
0							5.07	48.78	57.48	8.70	3.12	17.67	14.55	0.60	4.14	12.27	20.25	
120							0.00	1.86	5.17	3.31	1.35	23.69	22.34	0.85	5.29	11.14	19.61	

## 10% Initial Solids Level

YSI		YSI		Hauser				Hauser				Hauser					
Elapsed time (h)	Solids (%)	Glucose (g/L)	Glucose (g/L)	Ethanol (g/L)	Ethanol (g/L)	Xylose (g/L)	Cellobiose (g/L)	Monomeric Glucose (g/L)	Soluble Glucose <sup>1</sup> (g/L)	Oligomeric Glucose <sup>1,2</sup> (g/L)	Monomeric Xylose (g/L)	Soluble Xylose (g/L)	Oligomeric Xylose <sup>2</sup> (g/L)	Galactose (g/L)	Total Galactose (g/L)	Arabinose (g/L)	Total Arabinose (g/L)
0	10.28		24.30	1.02	1.01	3.67	3.37	25.76	30.13	4.37	2.08	10.39	8.31	0.37	1.93	6.12	10.65
22	7.98	0.08	0.00	16.95	16.69	1.82	0.00	1.03	3.02	1.99	0.91	11.73	10.82	0.39	2.33	5.63	9.75
46		0.09	0.00	18.05	17.61	1.02	0.00	0.84	2.55	1.71	0.50	11.34	10.84	0.28	2.19	5.45	9.59
68.5	7.99	0.10	0.00	18.68	18.50	0.84	0.00	0.79	2.38	1.59	0.37	11.23	10.86	0.30	2.18	5.14	9.46
91.5		0.07	0.00	18.63	18.70	0.82	0.00	0.76	2.52	1.76	0.31	11.31	11.00	0.33	2.07	4.99	9.09
120	8.01	0.07		18.54			0.00	0.75	2.31	1.56	0.31	10.96	10.65	0.42	1.93	4.83	8.62
Updated CAT task values																	
0							3.37	25.76	29.39	3.63	2.08	9.62	7.54	0.37	2.08	6.12	10.47
120							0.00	0.84	2.20	1.36	0.29	10.74	10.45	0.33	2.17	4.74	8.85

<sup>1</sup> Includes cellobiose

<sup>2</sup> Difference between soluble and monomeric values

Byproducts						
Mannose (g/L)	Total Mannose (g/L)	Succinic Acid (g/L)	Lactic Acid (g/L)	Glycerol (g/L)	Acetic Acid (g/L)	Xylitol (g/L)
0.00	0.09	0.00	3.54	0.21	1.95	
0.00	0.12	0.09	3.48	1.89	2.01	
0.00	0.09	0.11	3.47	2.10	2.08	
0.00	0.08	0.09	3.43	2.16	2.07	
0.00	0.08	0.18	2.93	1.86	1.85	1.67
0.00	0.08	0.22	2.89	1.87	1.87	2.21
0.00	0.06	0.00	3.48	0.30	2.01	0.00
0.00	0.00	0.34	3.05	2.88	2.83	2.21

Byproducts						
Mannose (g/L)	Total Mannose (g/L)	Succinic Acid (g/L)	Lactic Acid (g/L)	Glycerol (g/L)	Acetic Acid (g/L)	Xylitol (g/L)
0.00	0.31	0.01	2.86	0.15	1.06	
0.00	0.04	0.04	2.75	1.00	0.88	
0.00	0.04	0.06	2.64	0.97	0.98	
0.00	0.04	0.00	2.64	1.01	0.98	
0.00	0.04	0.11	2.10	0.85	0.90	1.74
0.00	0.04	0.29	2.44	1.03	1.16	1.90
0.00	0.00	0.01	2.86	0.15	1.06	0.00
0.00	0.00	0.63	1.87	1.11	1.16	1.90

Solids Analysis (% Dry weight)									
Glucose	Xylose	Galactose	Arabinose	Mannose	Klason Lignin	Acid Soluble Lignin	Total Ash	Total Solids	Insol Solids
34.04	24.42	1.30	7.30	0.25	13.78	7.77	0.92	15.65	16.69
12.81	22.01	3.81	7.41	0.24	19.65	9.95	1.95	17.6	7.23

Solids Analysis (% Dry weight)									
Glucose	Xylose	Galactose	Arabinose	Mannose	Klason Lignin	Acid Soluble Lignin	Total Ash	Total Solids	Insol Solids
36.00	21.65	1.42	7.63	0.29	13.02	8.05	1.04	14.19	4.66
10.65	15.79	2.77	4.53	2.43	20.12	10.83	1.54	11.75	2.51